Biomass and leaf-level gas exchange characteristics of three African savanna C_4 grass species under optimum growth conditions

K. B. Mantlana^{1,2,3}*, E. M. Veenendaal², A. Arneth⁴, V. Grispen², C. M. Bonyongo¹, I. G. Heitkonig² and J. Lloyd⁵

¹ Harry Oppenheimer Okavango Research Center (HOORC), University of Botswana, Private Bag 285, Maun, Botswana, ²Nature Conservation and Plant Ecology Group, Department of Environmental Sciences, Wageningen University and Research Centre, Borne steeg 69, NL 6708 PD, Wageningen, The Netherlands, ³Global Change and Biodiversity Programme, South African National Biodiversity Institute, Kirstenbosch Research Centre, P/Bag X 7, Cape Town, South Africa, ⁴Department of Physical Geography and Ecosystems Analysis, Lund University Sölvegatan 12, 223 62 Lund, Sweden and ⁵Earth and Biosphere Institute, School of Geography, University of Leeds, LS2 9JT Leeds, U.K.

Abstract

C4 savanna grass species, Digitaria eriantha, Eragrostis lehmanniana and Panicum repens, were grown under optimum growth conditions with the aim of characterizing their above- and below-ground biomass allocation and the response of their gas exchange to changes in light intensity, CO2 concentration and leaf-to-air vapour pressure deficit gradient (D1). Digitaria eriantha showed the largest above- and below-ground biomass, high efficiency in carbon gain under light-limiting conditions, high water use efficiency (WUE) and strong stomatal sensitivity to D₁ $(P = 0.002; r^2 = 0.5)$. Panicum repens had a high aboveground biomass and attained high light saturated photosynthetic rates (A_{sat}, 47 μmol m⁻² s⁻¹), stomatal conductance, (gsat, 0.25 mol m-2 s-1) at relatively high WUE. Eragrostis lehmanniana had almost half the biomass of other species, and had similar A_{sat} and g_{sat} but were attained at lower WUE than the other species. This species also showed the weakest stomatal response to Di $(P = 0.19, r^2 = 0.1)$. The potential ecological significance of the contrasting patterns of biomass allocation and variations in gas exchange parameters among the species are discussed.

Key words: above- and below-ground biomass, C₄ grasses, Digitaria eriantha, Eragrostis lehmanniana, gas exchange characteristics, leaf-to-air vapour pressure deficit, Okavango Delta, Panicum repens

Résumé

On a fait pousser des espèces herbeuses de savane de type C4, Digitaria eriantha, Eragrostis lehmanniana et Panicum repens, dans des conditions optimales dans le but de caractériser l'allocation de leur biomasse aérienne et racinaire et la réponse de leur échange gazeux à des changements d'intensité de la lumière, de concentrations de CO2, et à un gradient déficitaire (D1) de pression de vapeur feuille-air. D. eriantha montrait la plus grande biomasse aérienne et racinaire, une grande efficience de l'assimilation de carbone dans des conditions de luminosité limitée, une grande efficience d'utilisation de l'eau (WUE) et une forte sensibilité des stomates à $D_1(P = 0.002; r^2 = 0.5)$. P. repens avait une grande biomasse aérienne et atteignait des taux photosynthétiques élevés en lumière saturée (Assat, 47 μmol m⁻² s⁻¹), et une conductance stomatique (g_{sat} 0.25 mol m-2 s-1) à une WUE relativement élevée. E. lehmanniana avait une biomasse qui était presque la moitié de celle des autres espèces et avait un Asat et un gsat similaires mais qui étaient atteints à une WUE plus basse que les autres espèces. Cette espèce montrait aussi la plus faible réponse stomatique à D_1 (P = 0.19, $r^2 = 0.1$). L'on discute de la signification écologique potentielle de ces schémas contrastés d'allocations de biomasse et des variations des paramètres des échanges gazeux entre les espèces.

Introduction

The main constraints on plant and ecosystem productivity in the seasonally dry tropical savanna-wetland mosaic of

^{*}Correspondence: E-mail: mantlana@sanbi.org

the Okavango Delta in north-western Botswana are low soil water availability during the dry season, periodic flooding, generally low soil fertility, fire and grazing. The wet season (November-April) in this environment, when over 90% of the 400 mm annual rain falls, coincides with high leaf-to-air vapour pressure deficit (D; >4.5 kPa) and leaf temperatures (T1) may exceed 35°C.

A main factor that is supposed to regulate the adaptation of plants to environmental constraints is plasticity in the pattern of resource allocation (Lambers & Poorter, 1992). Plants that originate from infertile soils frequently display greater allocation of biomass to fine roots, and to increase the absorbing area (Baruch, 1994), invest less biomass to produce root length and produce fine roots with greater specific root length (SRL) (Lambers & Poorter, 1992). For grasses in the Okavango Delta, below ground accumulation of carbon and nutrient reserves would also allow for quick recovery of photosynthetic biomass after fires (Baruch, Ludlow & Davis, 1985). Previous studies have reported contrasting patterns in below- and aboveground biomass allocation in tropical and temperate grasses (Veenendaal, Shushu & Scurlock, 1993; Baruch, 1994; Ryser & Lambers, 1995), which suggest amplitude for resource partitioning.

C₄ species constitute a high proportion of the grass flora in the Okavango Delta. C4 photosynthesis involves at least three different biochemical pathways that differ in the mechanisms used to transport CO2 to the bundle sheath cells and also to regenerate phosphoenol pyruvate (Hatch, 1987). These biochemical pathways are NAD-malic enzyme species (NAD-ME), NADP-malic enzyme species (NADP-ME) and PEP carboxykinase species (PCK). Plants that use C4 metabolism possess a CO2 concentrating mechanism that enables them to have high nitrogen use efficiency, high water use efficiency, preference of high light intensities and high optimum leaf temperatures for CO2 uptake. Such adaptations make these plants particularly adapted for survival in the infertile and seasonally dry tropical ecosystems that often experience high vapour pressure deficits.

The aim of this work was to quantify the biomass allocation and to elucidate important information about the ecophysiology of little known, but commonly found, C4 grass species under optimum growth conditions. In the Okavango Delta, these species are found in different habitats: Digitaria eriantha Steud occurs in a wide-range of drier often partially shaded habitats in woodlands and islands and, Eragrostis lehmanniana Nees dominates dry, open and disturbed habitats, while Panicum repens L. dominates moist seasonally flooded areas of floodplains (Gibbs Russel et al., 1990). This was the first study to investigate gas exchange characteristics of C4 grasses from the Okavango Delta. Growing these plants under similar growth conditions (optimum water and nitrogen levels, high air temperature and high vapour pressure deficit) that mimic a period of high productivity with plant material sourced from their natural environment could provide insights into their potential responsiveness in natural conditions.

Materials and methods

Growth conditions

Vegetative cuttings of a tillering perennial, Digitaria eriantha Steud (NADP-ME), a tufted perennial Eragrostis lehmanniana Nees (NAD-ME), and a rhizomatous perennial Panicum repens L. (PCK), were collected from a floodplain in the Okavango Delta. Digitaria eriantha is native to Southern Africa. It is regarded as drought tolerant, and considered to have low shade tolerance, but high fire tolerance (Bogdan, 1977; Gibbs Russel et al., 1990). When collected from the field D. eriantha was growing under or very near to trees. Eragrostis lehmanniana is a seed-banking species and following fire, it re-germinates when soil water content becomes favourable. It dominates disturbed well-drained sandy soils (Gibbs Russel et al., 1990). When it was collected, E. lehmanniana was growing in open areas. Tropical and north Africa and the Mediterranean are the native range of P. repens but is now widespread throughout the tropics (Gibbs Russel et al., 1990). It favours open sunny conditions on moist, often sandy soil, and can withstand occasional flooding, but its rhizomes can stand prolong dry periods. Panicum repens was collected in the middle of a seasonal floodplain. Common characteristic among these species is that they are grazed extensively by herbivores (Gibbs Russel et al., 1990).

The plants were then planted into large bags (three cuttings per bag, 19 × 7.5 cm) to provide for ample rooting space, and kept outside a shade house to allow full exposure to light and ambient air temperatures which ranged from 35 to 39°C. The bags were filled with sandy soil from Lake Ngami and watered daily. After 5 weeks, the cuttings were transferred (one vegetative cutting of one species per bag) into new bags filled with uniform river sand. After transplanting, each plant received a fertilizer application. The fertilizer was applied as 10 ml liquid plant

food (Supranure, containing 11% N, 7.3% P and 3.7% K) diluted in 5 l of tap water. Each bag was given 200 ml of this solution. Fertilizer was re-applied after 3.5 weeks before the start of the photosynthesis measurements.

Harvest

After 8 weeks of growth and photosynthesis measurements had been completed, plants were harvested. The soil of roots of all individual plants was carefully rinsed with water, to minimize root damage or loss. Subsequently, each plant was separated into roots and shoots, and in the case of *P. repens*, also rhizomes. Subsamples of five leaves per shoot were taken to determine leaf area with a leaf area meter (model 3100 area meter; Li-Cor Inc., Lincoln, NE, U.S.A.). Subsamples of the roots were taken to determine SRL, using a ruler. All plant samples (roots and shoots) were oven-dried for 48 h at 80°C to determine dry weight. Total biomass, shoot—root ratio (SRR), leaf area ratio (LAR) and specific leaf area (SLA) were determined according to Beadle (1993). SRL was determined by dividing the length of the root sample by the dry weight.

Photosynthesis characteristics

Steady-state leaf gas exchange measurements were made using an open gas exchange system (LI-6400; Li-Cor Inc.) on youngest and fully expanded leaves from at least four randomly selected plants or separate tillers of each species. Measurements were made between 08.00 and 10.00 hours, to avoid midday stomatal depression. The leaf temperature inside the 6-cm2 leaf chamber was not controlled, but did not exceed 40°C, the temperature threshold above which C4 net photosynthetic assimilation may become inhibited (Lawlor, 2001). During the gas exchange measurements, all plants experienced similar leaf temperature and leaf-to-air vapour pressure deficit (D1) values (c. 37°C and 4.3 kPa respectively). Leaf areas were calculated using leaf dimensions of the enclosed leaf sections in the chamber (Anderson et al., 2001). Measurements were recorded only after the photosynthetic rate and stomatal conductance were considered constant and at equilibrium with the ambient conditions within the gas exchange cuvette.

During measurements both the leaf-to-air vapour pressure deficit, D_l , and leaf temperature, T_l , in the chamber did not always exactly match the ambient conditions because these two parameters are affected by leaf transpiration rate

(Beale, Morison & Long, 1999); T₁ typically exceeded air temperature at time of measurements by not more than 1-2°C. The red/blue LED light source inside the chamber allowed for automatic changes of light intensity (I) of 2000, 1500, 1000, 500, 200, 100, 50, 20 and $0 \mu \text{mol m}^{-2} \text{ s}^{-1}$. Apparent light use efficiency (ϕ) was estimated as the slope of the first four points in the linear part of the light response curve. The qualification, apparent, is used as the estimate is based on incident and not absorbed photon flux (Lawlor, 2001). Light-saturated photosynthetic rate, Asat, light-saturated stomatal conductance, gsat, the ratio of ambient to internal CO2 concentration ([CO2]), C1/Ca ratio, and water use efficiency, WUE, were obtained at high photon irradiance (>1500 µmol m⁻² s⁻¹) and at an ambient [CO₂] of 380 μmol mol⁻¹. WUE was determined as A_{sat}/g_{sat}.

CO2 response curves were then determined at high photon irradiance (2000 µmol m⁻² s⁻¹) and at different chamber [CO2] in the sequence ambient, 380, 300, 200, 100, 50, 400, 400, 600, 700 and 800 μmol mol⁻¹. The different CO2 concentrations were obtained by means of a portable CO2/air mixture tanks and automatically controlled by a CO2 injector. From each CO2 response curve, we determined the CO_2 compensation point, Γ , the initial slope, α, and the light and CO2 saturation rate of photosynthesis (A_{pot}) . Γ was determined by extrapolating the linear portion of the CO2 response curve to intercept the xaxis, and carboxylation efficiency (α; the initial slope of the CO2 response curve) was determined as the slope of the first four points in the linear part of the CO2 response curve (Polley et al., 1992). Gas phase limitation to photosynthesis, L_g, was estimated from the CO₂ response curves as $[(A_{pot} - A)/A_{pot}] \times 100$, (Farquhar & Sharkey, 1982; Ripley et al., 2007), where A is net photosynthesis at ambient chamber [CO₂], 380 µmol mol⁻¹. To test the significance of differences among the species in growth parameters and gas exchange parameters, data were analysed with univariate analysis of variance (ANOVA), using Tukev's HSD test. Statistical analyses were performed using the spss statistical package (SPSS 11.0 for Windows; SPSS Inc., Chicago, IL, USA).

Results

Plant growth

Digitaria eriantha and P. repens had almost twice as much biomass, dry weight, as that of E. lehmanniana. SLA realized

Species	C ₄ subtype	n	Biomass (g)	SRR (g g ⁻¹)	SRL (cm g ⁻¹)	$LAR~(cm^2~g^{-1})$	SLA (cm ² g ⁻¹)
Digitaria eriantha	NADP-ME	8	39.4 ^b	3.8 ^b	326 ^b	90°	245 ^b
Eragrostis lehmanniana	NAD-ME	11	21.2a	3.7 ^b	211a	36ª	191ª
Panicum revens	PCK	12	40.0 ^b	1.8a	170 ^a	63 ^b	191ª

Table 1 C4 photosynthetic subtypes and mean values and standard deviations of biomass allocation characteristics of the three savanna grass species

SRR, shoot-root ratio; SRL, specific root length; LAR, leaf area ratio; SLA, specific leaf area Different superscripts in each column indicate significant differences at P = 0.05; Tukey HSD test.

with this biomass was highest in D. eriantha and similar between the other two species (Table 1). As was the case with biomass, the ratio of leaf area to total plant weight (LAR) of D. eriantha and P. repens was significantly higher (P < 0.05) than that of E. lehmanniana. Root length per unit root biomass (SRL) of D. eriantha was significantly higher (P < 0.05) than that of the other two species. Digitaria eriantha had similar SRR with E. lehmanniana that were almost double that of P. repens (Table 1).

Photosynthetic characteristics

Typical of C4 species, net photosynthesis in all the species was not light saturated (Fig. 1). Digitaria eriantha had highest mean apparent light use efficiency $[\phi; 0.067 \text{ mol}]$ (CO2) mol-1 (incident photon)] which was significantly higher (P = 0.021) than that of P. repens [0.056 mol (CO_2) mol-1 (incident photon); Table 2].

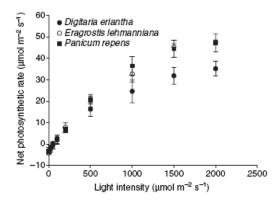


Fig 1 Mean values and standard deviations of the response of net CO₂ assimilation to photon irradiance in leaves of three C₄ grasses. Each datum point is a mean of at least four measurements. For clarity of the figure, only standard deviations for A are shown

Eragrostis lehmanniana and P. repens displayed similar mean A_{sat} and mean g_{sat} values (47 μ mol m⁻² s⁻¹ and 0.27 mol m⁻² s⁻¹ respectively) that were significantly higher (P < 0.05) than in D. eriantha (Table 3). Mean WUE values were similar between D. eriantha and P. repens (c. 190 µmol CO2 mol-1 H2O) and were significantly higher (P < 0.05) than those of E. lehmanniana (Table 3). The rank order of the species remained the same even when WUE was determined as A/E (net photosynthesis/transpiration) (data not shown). Digitaria eriantha and P. repens displayed similar mean Ci/Ca ratio (0.11 and 0.12 respectively) that were significantly lower (P < 0.05) than that of E. lehmanniana (0.18; Table 3). Consistent with these observations, there were similar slopes of the relationship between A_{sat} and g_{sat} in D. eriantha and P. repens (Fig. 2). This relationship was linear and close to proportional in D. eriantha and P. repens, in contrast to that of E. lehmanniana which was nonlinear. In E. lehmanniana, $A_{\rm sat}$ increased with $g_{\rm sat}$ until values of c. 0.3 mol m⁻² s⁻¹, above which a further stomatal opening did not lead to any evident further increase in A_{sat} .

Despite considerable scatter in the data, $g_{\rm sat}$ and $C_{\rm i}/C_{\rm a}$ ratio decreased with an increase in D1 in all the species (Fig. 3). The strongest and significant linear correlations between g_{sat} versus D_l and C_i/C_a ratio versus D_l relationships were found in P. repens $(P < 0.05, r^2 = 0.5 \text{ and } 0.6)$ respectively). In contrast, both relationships were weak and nonsignificant (P > 0.05, $r^2 = 0.1$) in E. lehmanniana. In all the species, the relationship between A_{sat} and D_l followed a similar pattern to that of g_{sat} versus D_{l} . (data not

Consistent with C4 photosynthesis, CO2 assimilation rates increased rapidly, in response to increasing internal CO_2 concentrations, C_i , up to C_i values of 50-80 μ mol mol⁻¹, before becoming saturated at high chamber [CO₂] (Fig. 4). Also, typical of C4 plants, CO2 compensation points of all species were low, c. 6 µmol m⁻² s⁻¹, with no significant differences (P > 0.05) between the species

Table 2 Mean values and standard deviations of carboxylation efficiency (CE), CO_2 compensation point (Γ), light use efficiency (\varnothing), dark respiration (R_d), gas phase limitation to photosynthesis (L_z) and leaf-to-air vapour pressure defict (D_l) for the three grass species

	CE $[\mu \text{mol m}^{-2} \text{ s}^{-1}]$ $(\mu \text{mol mol}^{-1})^{-1}$	$\Gamma (\mu \text{mol m}^{-2} \text{ s}^{-1})$	ø (mol mol ⁻¹)	Lg (%)
Digitaria eriantha	2.19 (0.84) ^a	6.96 (1.19) ^a	0.067 (0.01) ^b	22.14 (4.44) ^b
Eragrostis lehmanniana	1.66 (0.33) ^a	5.98 (3.34) ^a	0.063 (0.01) ^{ab}	14.44 (3.06) ^a
Panicum repens	2.59 (1.08) ^a	6.91 (1.31) ^a	0.056 (0.02) ^a	17.47 (2.39) ^{ab}

Different superscripts in each column indicate significant differences at P = 0.05; Tukey HSD test.

Table 3 Mean values and standard deviations of light saturated net photosynthetic rate $(A_{\rm sat})$, light and ${\rm CO_2}$ saturated net photosynthetic rate $(A_{\rm pot})$, light saturated stomatal conductance $(g_{\rm sat})$, the ratio between ambient and intercellular ${\rm CO_2}$ (C_t/C_a) and water use efficiency (WUE) for the three species at saturating light $(1600-2000~\mu{\rm mol}~{\rm m}^{-2}~{\rm s}^{-1})$ and at ambient $[{\rm CO_2}]$ of 380 $\mu{\rm mol}~{\rm mol}^{-1}$

	$A_{\rm sat}(\mu{\rm mol~m^{-2}~s^{-1}})$	$A_{\rm pot}~(\mu{\rm mol~m^{-2}~s^{-1}})$	$g_{\rm sat}~({\rm mol~m^{-2}~s^{-1}})$	C_i/C_a	WUE (μ mol CO $_2$ mol $^{-1}$ H $_2$ O)
Digitaria eriantha	35.28 (6.76) ^a	41.23 (4.37) ^a	0.19 (0.05) ^a	$0.11 (0.04)^{a}$	190.3 (14.2) ^b
Eragrostis lehmanniana	47.47 (4.04) ^b	52.33 (2.25) ^b	0.31 (0.08) ^b	$0.18 (0.05)^{b}$	161.8 (28.6) ^a
Panicum repens	47.33 (6.86) ^b	51.30 (8.07) ^b	0.26 (0.05) ^b	$0.12 (0.03)^{a}$	184.2 (12.9) ^b

Different superscripts in each column indicate significant differences at P = 0.05; Tukey HSD test.

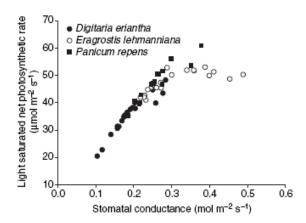


Fig 2 Relationship between light saturated net CO_2 assimilation and stomatal conductance of leaves of C_4 species. Each point represents an individual measurement at saturating light (1600–2000 μ mol m⁻² s⁻¹) and at ambient [CO₂] of 380 μ mol mol⁻¹

(Table 2). Initial slopes of the CO_2 response curves (α), showed no significant differences (P=0.306) between the species, ranging from 1.7 to 2.6 μ mol m⁻² s⁻¹ (μ mol - mol⁻¹)⁻¹ (Table 2).

In *D. eriantha* and *P. repens*, raising chamber C_a values above ambient [CO₂] led to a relatively small increase in C_i , up to 120 μ mol mol⁻¹ (Fig. 4). In contrast, at high C_a , C_i of *E. lehmanniana* increased to almost 400 μ mol mol⁻¹. Similar to A_{sat} , highest mean light and CO₂ saturated photo-

synthetic rates (A_{pot}) values were found in *E. lehmanniana* and *P. repens* (c. 52 μ mol m⁻² s⁻¹) and were significantly higher (P < 0.05) than that found in *D. eriantha* (41 μ mol m⁻² s⁻¹). Mean gas phase limitation (L_g) values, which indicate the magnitude of increase of photosynthesis at ambient CO₂ concentration relative to A_{pot} , were highest in *D. eriantha* (22%) while *E. lehmanniana* had lowest values (14%; Table 2).

Discussion

The aim of this study was to characterize plant biomass and leaf-level gas exchange of commonly found C_4 grasses with the aim of improving our knowledge in predicting their field performance in natural environments. In the long term, our objective was to identify representative species that may be used to gather information that would be useful when constructing the carbon budget in the ecosystems of the Okavango Delta.

Although caution is necessary, when comparing allocation patterns in plants when only single harvests are taken (Lambers & Poorter, 1992), the long growth period and the similar order of magnitude of the final biomass of the plants at the end of the experiment suggest that some differences can still be considered to reflect true differences allocation patterns. For instance, the larger biomass in D. eriantha was associated with high SRL. Increased SRL

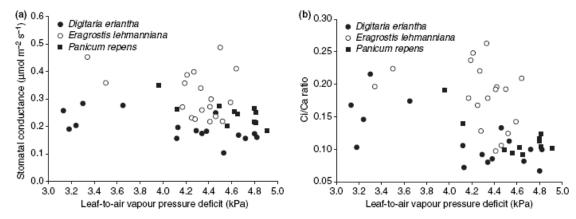


Fig 3 The response of stomatal conductance (a) and C/Ca ratio (b) to leaf to air vapour pressure deficit of the three species. Each point represents an individual measurement at saturating light (1600-2000 μmol m⁻² s⁻¹) and at ambient [CO₂] of 380 μmol mol⁻¹

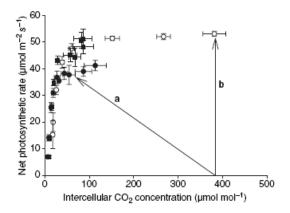


Fig 4 Mean values and standard errors of the response of net CO2 assimilation to CO2 in leaves of three C4 grasses. Each datum point is a mean of at least four measurements. The vertical line, denoted b, represents photosynthetic rate at ambient [CO2] if resistance to diffusion was zero and that denoted a, represents the limitation on net photosynthesis imposed by CO2 diffusion. Line a has a slope set by the stomatal conductance and both lines intercept the C1 axis at ambient [CO2]. Symbols are the same as those in Fig. 1

results from longer root length (and hence larger root surface area) per unit carbon invested (Eissenstat, 1991), and indicates potential for high rates of root growth in favourable soil conditions. Further, the high LAR displayed by D. eriantha, which was probably due to its high SLA, suggests a physiological trait to optimize photon harvesting. The advantage of a high SLA is a higher photosynthetic rate per unit leaf weight (Roderick & Cochrane,

2002). Indeed, our results showed that despite lowest area based photosynthetic rates in D. eriantha, mass based photosynthetic rates between the species were similar. Taken together, the above- and below-ground allocation of biomass in D. eriantha suggests high potential of this species to exploit the seasonal pulses in nutrient and water availability and to tolerate different levels of shading that it encounters under natural conditions, hence its occurrence in a wide range of habitats within the Okavango Delta and throughout Southern Africa (Gibbs Russel et al., 1990).

The low SRR found in P. repens indicates a strong priority for shoot growth. A trait particularly important in wetlands where under more stable moisture conditions shoot competition may be more important. The high productivity of P repens makes it an important grazing resource for game and domestic livestock (Gibbs Russel et al., 1990; Van Bommel et al., 2006). However, the small root allocation in this species could imply low capacity to tolerate dry conditions and also reduced capacity for soil water and nutrient uptake when growing under natural conditions.

The range of mean A_{sat} and g_{sat} , 35–48 μ mol m⁻² s⁻¹, 0.19-0.31 mol m⁻² s⁻¹, respectively, observed in this study was comparable to that reported for other C4 species optimum grown under conditions (25 60 μ mol m⁻² s⁻¹ and 0.2-0.4 mol m⁻² s⁻¹ respectively) (Lawlor, 2001). The positive curvilinear relationship between g_{sat} and A_{sat} found in E. lehmanniana implies more stress on g_{sat} than on photosynthetic capacity (Hetherington & Woodward, 2003), while the close to proportional and almost linear relationship in D. eriantha and P. repens, if maintained would result in a constant Ci (Jones, 1987). However, this relationship did not pass through the origin; therefore C_i could not be constant.

The range of mean C_i/C_a values of these species, 0.11– 0.18, was on the lower side of those previously reported at moderate vapour pressure deficits and well-watered C4 species (0.2-0.4; Lawlor, 2001). Such low Ci/Ca values were probably a reflection of both the direct response of stomata to high D and also the relatively high photosynthetic capacity (Condon et al., 2002). The Ci/Ca ratio is determined by the balance between the supply of CO2 to the leaf interior and the demand for CO2 (Farquhar & Sharkey, 1982). The low Ci/Ca ratio and high WUE displayed by D. eriantha and P. repens indicate that carbon gain occurred at minimal loss of water. However, our results suggest that the low Ci/Ca ratio in P. repens might have been due to high photosynthetic capacity as indicated by its high A_{sat} , in contrast, the low C_i/C_a ratio of D. eriantha might have been a result of lower stomatal conductance, as indicated by the low A_{sat} .

The slope of the relationship between g_{sat} and D_l has been used in C4 species to indicate stomatal sensitivity to D_I (Bunce, 1983; Kawamitsu, Yoda & Agata, 1993; Maroco, Pereira & Chaves, 1997). The lower sensitivity of g_{sat} to D_l and the weaker response of A_{sat} to D_l displayed by E. lehmanniana suggest better adaptation to seasonally dry environments where high D_1 is associated with the wet season. High stomatal sensitivity, as found in P. repens and D. eriantha might be ecologically disadvantageous, since plants might miss the opportunity for photosynthesis when there is available soil water to meet the transpirational demand. A study on the three Sahelian C4 grass species attributed pronounced stomatal sensivity to drought resistance, while a lack of stomatal sensitivity was associated with drought escaping species (Maroco et al., 1997).

Well-established theory suggests that, on average, NADP-ME species have higher ϕ than NAD-ME species with PCK species being intermediate (Ehleringer & Pearcy, 1983). This pattern was not apparent in this study and may have been masked by the use of only one species per C₄ subtype. However, the high ϕ found in D. eriantha indicates higher efficiency of leaf photosynthesis in light-limiting conditions (Ehleringer & Pearcy, 1983). Functionally, this may be a useful trait as this species is sometimes found in savanna communities with higher leaf area indices (Gibbs Russel et al., 1990). Because of the presence of a CO₂-concentrating mechanism, photorespiration is suppressed and photosynthesis

operates at near CO_2 -saturation in well-watered C_4 plants (Von Caemmerer, 2000). Consistent with this theory, our results showed Γ of <10 μ mol mol⁻¹ and a relatively small increase, 14–22%, in photosynthesis at short-term exposure to high [CO_2].

In conclusion, substantial differences in above- and below-ground biomass allocation may potentially contribute to their different abilities to extract water and nutrients, and probably reflect the adaptations to the different habitats they dominate. Digitaria eriantha produced high above- and below-ground biomass, showed high WUE and high adaptation to low light conditions. If this pattern holds in the field, it would suggest that both morphological (biomass accumulation) and physiological (gas exchange characteristics) play a role in facilitating this species relative abundance in a wide range of habitats within the savanna ecosystem. Gas exchange parameters in E. lehmanniana showed the weakest response to high evaporation demand, indicative of better adaptation to seasonally dry environments and consistent with the dry, open and disturbed habitats dominated by this species. The high above ground biomass allocation of P. repens may be indicative of a mechanism to cope with herbivory. On the other hand, high stomatal sensitivity suggests a physiological basis for this species greater abundance in wet areas of the Okavango Delta.

The experimental design of this study prevented the detection of plant-plant interactions which would be found under natural conditions. Moreover, as these plants were grown under ambient [CO2] and exposed only for short periods to high CO₂ chamber environment, it was not possible to compare our results to other C₄ grasses grown under high [CO₂] (e.g. Roumet & Roy, 1996). A more complete comparative assessment of the morphological and leaf-level photosynthetic characteristics of these species awaits the incorporation of species-specific leaf nitrogen content. We suggest that the adaptive traits observed here could be tested over a larger number of species and growing them over a longer period of time, combined with manipulation of water and/or fertilization regimes, to allow for a further identification of evolutionary pressures that best reflect selection for functionally adaptive morphological and physiological characteristics. Taken together, our results make a significant contribution to the understanding needed to predict the spatio-temporal distribution of these species and may also be useful in determining gross primary productivity of this savanna-wetland mosaic.

Acknowledgements

This research was funded by the Harry Oppenheimer Okavango Research Centre, University of Botswana, Wageningen University and the Max Planck Institute for Biogeochemistry. KBM also acknowledges a scholarship from the National Research Fund (South Africa). The authors are grateful to Dr Baruch (University Simon Bolsvar, Caracas, Venezuela) for comments and suggestions on this manuscript.

References

- Anderson, I.J.H., Maherali, H., Johnson, H.B., Polley, H.W. & Jackson, R.B. (2001) Gas exchange and photosynthetic acclimation over subambient to elevated CO2 in a C3-C4 grassland. Glob. Chang. Biol. 7, 693-707.
- BARUCH, Z. (1994) Responses to drought and flooding in tropical forage grasses: I. - production and allocation of biomass, leaf growth and mineral nutrients. Plant Soil 164, 87-96.
- BARUCH, Z., LUDLOW, M.M. & DAVIS, R. (1985) Photosynthetic responses of native and introduced C4 grasses from Venezuelan savannas. Oecologia 67, 338-393.
- BEADLE, C.L. (1993) Growth analysis. In: Photosynthesis and Production in a Changing Environment (Eds D. O. HALL, J. M. O. SCURLOCK, H. R. BOLHAR-NORDENKAMPF, R. C. LIEGOOD and S. P. LONG). Chapman and Hall, London.
- BEALE, C.V., MORISON, J.I.L. & LONG, S.P. (1999) Water use efficiency of C4 perennial grasses in a temperate climate. Agric. For. Meteorol. 96, 103-115.
- BOGDAN, A.V. (1977) Tropical Pasture and Fodder Plants (Grasses and Legumes). Longman, London and New York.
- Bunce, J.A. (1983) Differential sensitivity to humidity of daily photosynthesis in the field in C₃ and C₄ species. Oecologia 57, 262-265.
- CONDON, A.G., RICHARDS, R.A., REBETZKE, G.J. & FARQUHAR, G.D. (2002) Improving intrinsic water-use efficiency and crop yield. Crop Sci. 42, 122-131.
- EHLERINGER, J.R. & PEARCY, R.W. (1983) Variations in quantum yields among C3 and C4 plants. Plant Physiol. 73, 555-559.
- EISSENSTAT, D.M. (1991) On the relationship between specific root length and the rate of root proliferation; a field study using citrus rootstocks. New Phytol. 118, 63-68.
- FARQUHAR, G.D. & SHARKEY, T.D. (1982) Stomatal conductance and photosynthesis. Ann. Rev. Plant Physiol. 33, 317-346.
- GIBBS RUSSEL, G.E., WATSON, L., KOEKEMOER, M., SMOOK, L., BARKER, N.P., Anderson, H.M. & Dallwitz, M.J. (1990) Grasses of Southern Africa. Botanical Research Institute, Pretoria, South Africa.
- HATCH, M.D. (1987) C4 photosynthesis a unique blend of modified biochemistry, anatomy and ultrastructure. Biochim. Biophys. Acta 895, 81-106.

- HETHERINGTON, A.M. & WOODWARD, F.I. (2003) The role of stomata in sensing and driving environmental change. Nature 424,
- JONES, M.B. (1987) The photosynthetic characteristics of papyrus in a tropical swamp. Oecologia 71, 355-359.
- KAWAMITSU, Y., YODA, S. & AGATA, W. (1993) Humidity pretreatment affects the response of stomata and CO2 assimilation to vapour pressure difference in C3 and C4 plants. Plant Cell Physiol. 34, 113-119.
- Lambers, H. & Poorter, H. (1992) Inherent variation in growth rate between higher plants: a search for physiological causes and ecological consequences, Adv. Ecol. Res. 23, 87-261.
- LAWLOR, D.W. (2001) Photosynthesis, 3rd edn. Bios. Scientific Publishers, Oxford.
- MAROCO, J.P., PEREIRA, J.S. & CHAVES, M.M. (1997) Stomatal responses to leaf-to-air vapour pressure deficit in Sahelian species. Aust. J. Plant Physiol. 24, 381-387.
- POLIEY, H.W., NORMAN, J.M., ARKERBAUER, T.J., WALTER-SHEA, E.A., Greegor, D.H. & Bramer, B. (1992) Leaf gas exchange of Andropogon gerardii Vitman, Panicum virgatum L., and Sorghastrum nutans (L.) Nash in a tallgrass prairie. J. Geophys. Res. 97, 837-
- RIPLEY, B.S., GILBERT, M.E., IBRAHIM, D.G. & OSBORNE, C.P. (2007) Drought constraints on C4 photosynthesis: stomatal and metabolic limitations in C3 and C4 subspecies of Alloteropsis semialata. J. Exp. Bot. 58, 1351-1363.
- RODERICK, M.L. & COCHRANE, M.J. (2002) On the conservative nature of the leaf mass-area relationship. Anal. Bot. 89, 537-542.
- ROUMET, C. & ROY, J. (1996) Prediction of the growth response to elevated CO2: a search for physiological criteria in closely related grass species. New Phytol. 134, 615-621.
- Ryser, P. & Lambers, H. (1995) Root and leaf attributes accounting for the performance of fast- and slow-growing grasses at different nutrient supply. Plant Soil 170, 251-265.
- VAN BOMMEL, F.P.J., HETKONIG, I.M.A., EPEMA, G.F., RINGROSE, S., BONYONGO, C. & VEENENDAAL, E.M. (2006) Remotely sensed habitat indicators for predicting distribution of impala (Aepyceros melampus) in the Okavango Delta, Botswana. J. Trop. Ecol.
- VEENENDAAL, E.M., SHUSHU, D.D. & SCURLOCK, J.M.O. (1993) Responses to shading of seedlings of savanna grasses (with different C4 photosynthetic pathways) in Botswana. J. Trop. Ecol. 9, 213-229.
- Von Caemmerer, S. (2000) Biochemical Models of Leaf Photosynthesis. CSIRO Publishing, Collingwood, Australia.

(Manuscript accepted 16 January 2008)

doi: 10.1111/j.1365-2028.2008.00961.x